



Monitoring the effects of neonicotinoid pesticides on the phyllosphere and soil bacterial communities in a three-year soybean and corn rotation


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OBJECTIVES


 Characterize soybean and corn phyllosphere and soil bacterial community composition


 Determine the bacterial variation and temporal dynamics


 In a three-year soybean/corn crop rotation


 In response to neonicotinoid (neonic) seed treatment


METHODS


 Collecting samples


 **Phyllosphere:** 50-100g per sample of the plants’ middle leaves (144 samples in total) (see Table 2)


 **Soil:** 400-500g per sample of **bulk soil** from the upper 12-15 cm layer of soil and at 10 cm from the sampled plants (192 samples in total) (see Table 2)

 Extracting bacterial DNA, using PowerSoil DNA isolation kit


 Amplifying the hypervariable V5-V6 regions of 16S rRNA gene, using chloroplast-excluding 799F-1115R primers


 Sequencing DNA amplicons, using Illumina MiSeq


 Processing data into an amplicon sequence variants (ASVs) table for each sample and assigning taxonomy, using DADA2


 Analyzing data using R packages


RESULTS

 Plant host species explained 12% (phyllosphere) and 2% (soil) of the bacterial diversity.


 Host growth stage (= month in figure 2) was the strongest driver of the **phyllosphere** bacterial composition diversity and explained **more than 20%** of the soybean and corn phyllosphere bacterial variation.


 Year also explained the bacterial diversity in **soybean phyllosphere (11%)** and **soil (5%)**.



 **Neonic** had an impact on the phyllosphere and especially soil bacterial community composition, **individually and in interaction with host species, host growth stage and time (month and year)**.

 The effects of **neonic** on soil bacteria **increased during growing season** (July and August) and then **decreased** towards the end of the season (September).

CONCLUSIONS

 Plant host (species and growth stages) and time (month and year) are **stronger drivers** of variation in phyllosphere bacterial composition than neonic application; however, **Neonicotinoids in interaction** with this parameters influence the **phyllosphere** bacterial diversity.

 **Neonicotinoid pesticides** have an impact on the **soil** bacterial taxonomic composition. These impacts change over time (months and years) and are more significant in the **middle of the growing season**.



Neonicotinoid pesticides in interaction with **plant host** (species and growth stage) and **time** (month and year) influence **phyllosphere** and **soil bacterial diversity**.

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Neonicotinoids have an impact on the **soil taxonomic composition**, especially in the **middle of the growing season**.


Phyllosphere			Soil	
Variables	R2(%)	Pr(>F)	R2(%)	Pr(>F)
Host	12	0.001	2	0.001
Year	8	0.001	3	0.001
Month/Growth stage	14	0.001	3	0.001
Neonic	1	0.003	3	0.001
Host:Neonic	1	0.001	1	0.004
Year:Neonic	1	0.010	1	0.006
Month:Neonic	1	0.029	2	0.031
Host:Month:Neonic	1	0.019	-	-
Year:Month:Neonic	1	0.019	-	-

Soybean			Soil	
Variables	R2(%)	Pr(>F)	R2(%)	Pr(>F)
Year	11	0.001	5	0.001
Month/Growth stage	22	0.001	5	0.001
Neonic	1	0.023	4	0.001
Year:Neonic	1	0.033	2	0.005

Corn			Soil	
Variables	R2(%)	Pr(>F)	R2(%)	Pr(>F)
Month/Growth stage	27	0.001	9	0.001
Neonic	6	0.001	4	0.001

Table 1: Bacterial community taxonomic composition variation of both soybean and corn soil and phyllosphere (blue), as well as soybean (green) and corn (yellow) soil and phyllosphere, separately (PERMANOVA Bray-Curtis dissimilarities).

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STUDY DESIGN

Crop	Year	Habitat	Sampling Time		Number of Treatments	Replicate	Sample/ Replicate	Total Samples	Table 2: Sampling chart.	
			Month	Stage						
Soybean	2016	Leaf	July	Full Flowering	2	4	2	48/year		
			Aug.	Seeding						
			Sep.	Maturity						
	2018	Soil	May	Planting	2	4	2	64/year		
			July	Full Flowering						
			Aug.	Seeding						
Corn	2017	Leaf	July	Tasseling	2	4	2	48		
			Aug.	Blister						
			Sep.	Maturity						
	2018	Soil	May	Planting	2	4	2	64		
			July	Tasseling						
			Aug.	Blister						
			Sep.	Maturity						

WANT MORE DETAILS?

Figure 2: Bray-Curtis based non-metric multidimensional scaling (NMDS) plot of the phyllosphere (top) and soil (bottom) bacterial taxonomic composition show that host species (especially for phyllosphere) and time (month and year) are the main drivers of phyllosphere and soil bacterial diversity and explain it better than neonic treatment. However, the effects of neonic on soil bacterial composition are in interaction with time (especially month). These effects increase in the middle of growing season (July and August) and decrease towards the end of the season. Ellipses represent 99% confidence level.

Figure 3: The soybean and corn soil bacterial genera that are significantly associated with neonic treatment. The genera on the top of the graph (log2FoldChange > 0) are related to the non-treated samples, while the others (log2FoldChange < 0) are associated with the neonic-treated samples.